



9. Methods in Immunology

by Claudine Neyen and Bruno Lemaitre,

Ecole Polytechnique Fédérale de Lausanne

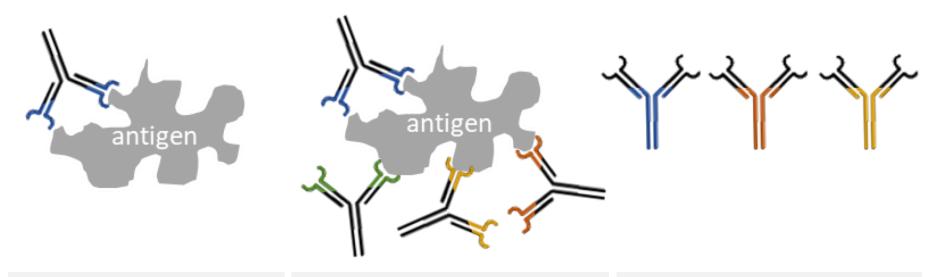
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Outline

- 9.1 Antibody-based tools
- 9.2 Identifying & quantifying antigens: **ELISA and Western Blot**
- 9.3 Cell-based assays to assess immune reactions: cytotoxicity and proliferation
- 9.4 Identifying, characterizing and purifying cell types: Flow cytometry and FACS

Antibodies as tools to identify molecules

- Antibodies (predominantly IgGs) are used for a variety of applications in research, diagnostics and therapy.
- Antibodies can be polyclonal (recognize many epitopes of the same antigen) or monoclonal (recognize a single epitope of the same antigen)
- Antibodies can be raised in different host species (e.g. mouse, rabbit, donkey, chicken, goat)



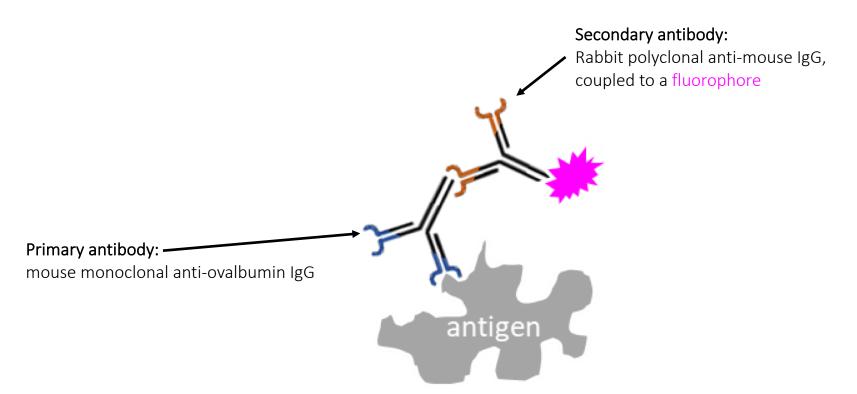
Monoclonal antibody

Polyclonal antibody

Antibodies from different species have species-specific Fc regions

Antibodies as tools to identify molecules

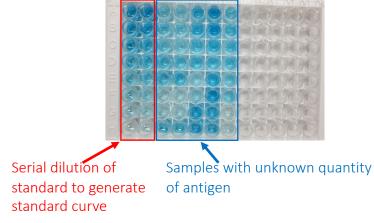
- Primary antibodies recognize a given antigen (e.g. mouse anti-ovalbumin IgG)
- Secondary antibodies recognize the Fc portion of an antibody from a different species (e.g. goat anti-mouse IgG)
- Antibodies can be **coupled to fluorescent molecules** (e.g. FITC, PE) **or to effector molecules** (e.g. enzymes like peroxidase)



ELISA (Enzyme-Linked ImmunoSorbent Assay)

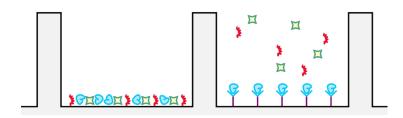
| What is it? | Microplate-based assay |
|----------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| What can it do? | Detect and quantify molecules (peptides, proteins, hormones,) from complex mixtures (e.g. serum, cell culture supernatant) |
| How does it work? | Antigen is immobilized on a solid surface An antigen-specific antibody is added to detect the antigen The enzyme coupled to the antibody catalyses a coloured substrate (dye) reaction that can be quantified in a spectrophotometer |
| \\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\ | |

What does an ELISA look like?

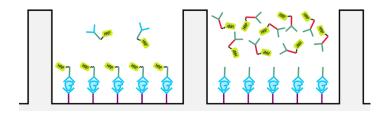


ELISA (Enzyme-Linked ImmunoSorbent Assay)

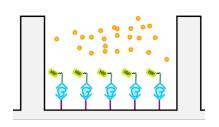
 Direct versus indirect capture ('sandwich ELISA')

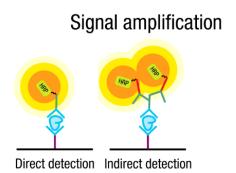


2. Direct versus indirect detection

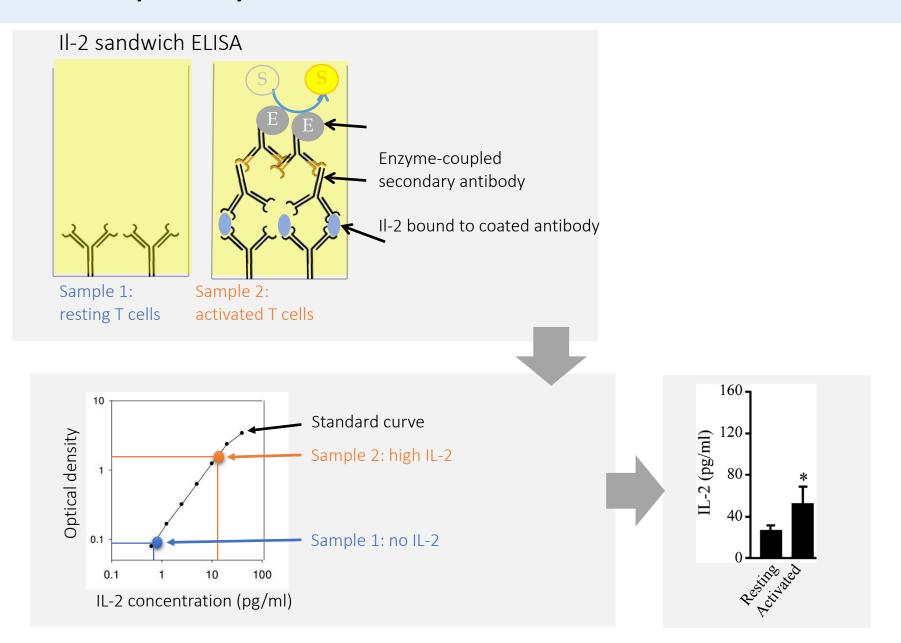


3. Substrate reaction





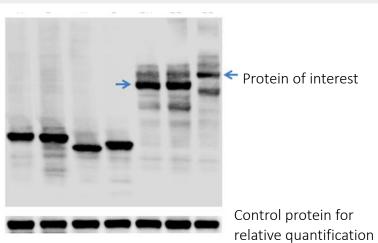
Example: Cytokine levels in serum or tissue culture



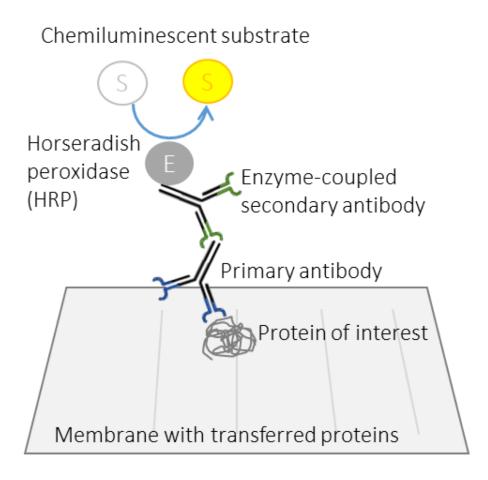
Western Blot

| What is it? | Solid-phase protein detection assay |
|-------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| What can it do? | Detect, quantify and characterize proteins and post-translational modifications (e.g. phosphorylation) |
| How does it work? | A protein mix is separated by gel electrophoresis and transferred to a nitrocellulose membrane (blot) An antigen-specific antibody is added to detect the antigen The enzyme coupled to the antibody catalyses a chemiluminescent reaction that can be visualized on film |

What does a Western Blot look like?

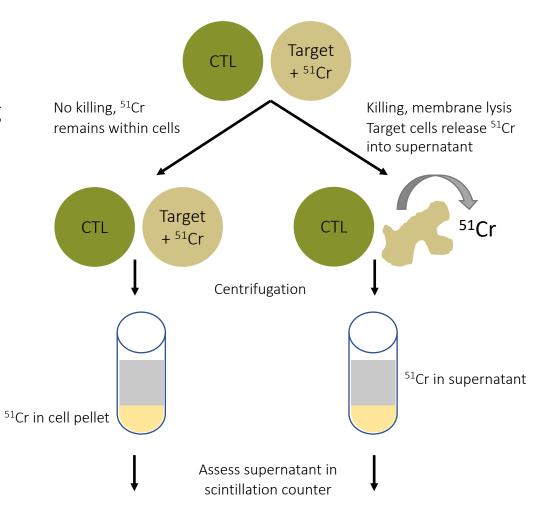


Western Blot



Cell-based assays: Cr51 release assay to assess cytotoxicity

- Used to assess CTL or NK cell function
- CTLs or NK cells kill by releasing granzyme and perforin, which disrupts target cell membrane
- Target cells are loaded with radioactive chromium



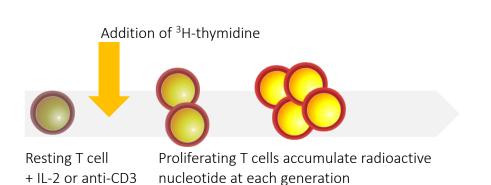
Cell-based assays: proliferation

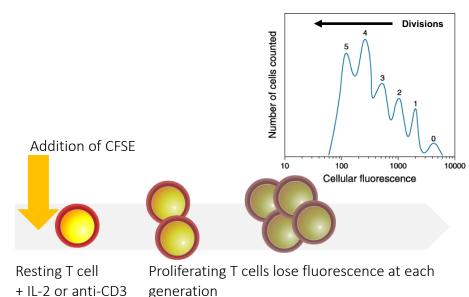
Radioactive labelling

- Cells are stimulated to induce proliferation
- Radiolabelled nucleotides are added to the medium
- Cells incorporate ³H-thymidine into their DNA during each round of replication
- After several days, cells are harvested, lysed, and radioactivity is counted

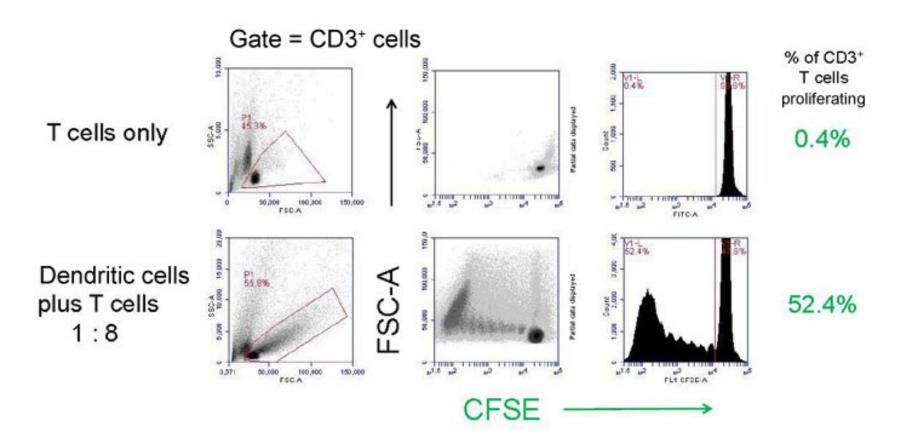
Fluorescent labelling

- Cells are labelled with a cytoplasmic fluorophore (e.g. CFSE)
- Cells are stimulated to induce proliferation
- At each division, daughter cells receive half the amount of the fluorophore
- After several days, cells are fixed and their fluorescence measured by flow cytometry





Example: T cell proliferation assay

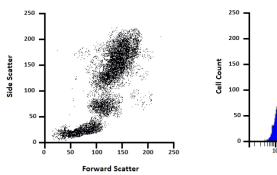


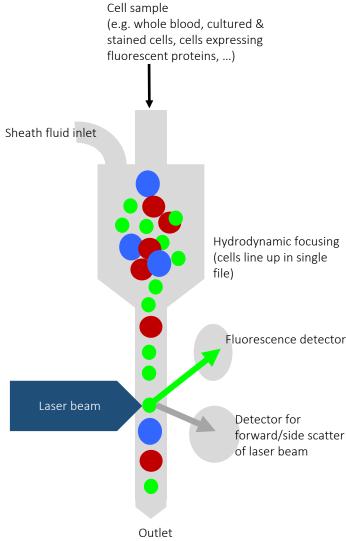
Flow cytometry: principle

Developed in 1954 by Walter Coulter

| What is it? | Laser-based single-cell fluidics technology |
|-------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| What can it do? | Characterize cells or particles |
| How does it work? | A single-cell suspension is passed in front of a laser Detectors measure how each cell scatters the laser beam fluorescence emitted from each cell |
| What does flow | ²⁵⁰ 7 |

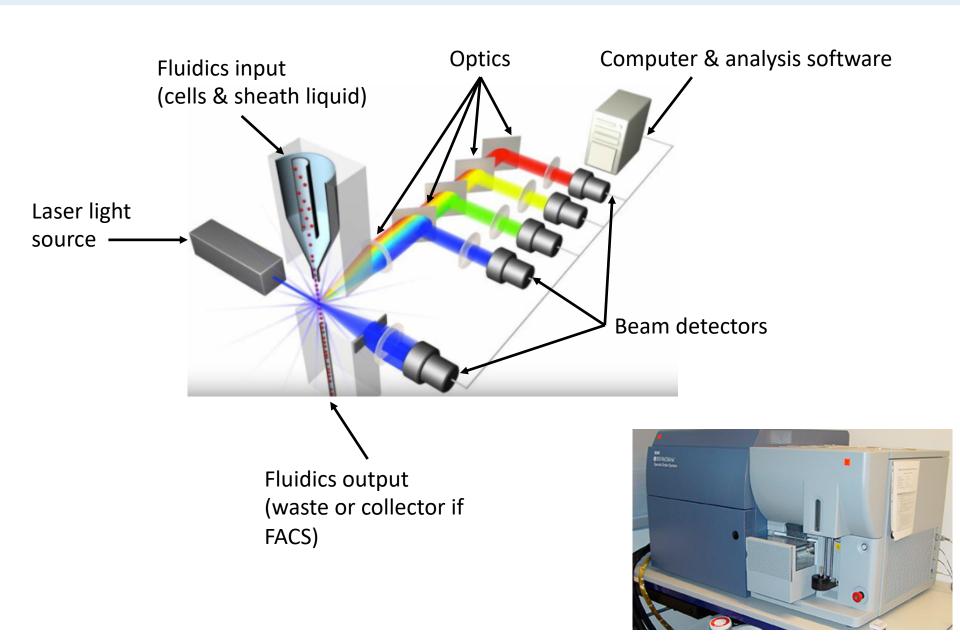
What does flow cytometry data look like?





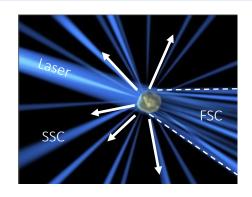
cytometry principles here

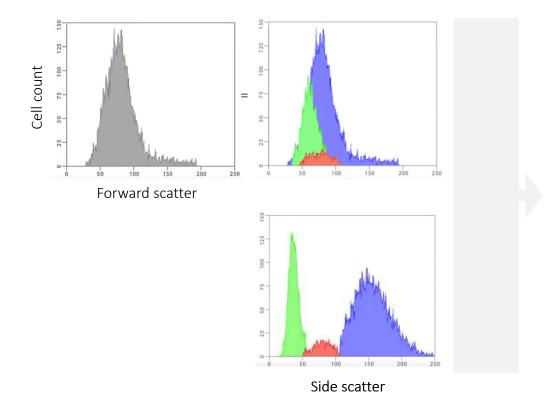
Flow cytometry: equipment



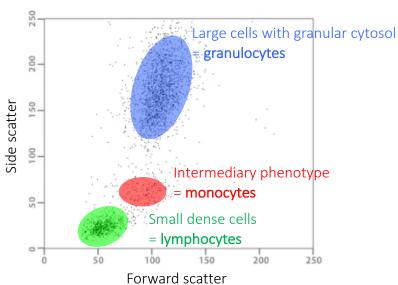
Flow cytometry with unstained cells

- A cell passing through the laser scatters the beam
 - Forward scatter (FSC) depends on cell size
 - Side scatter (SSC) depends on cell granularity
- Each event is counted and plotted on a scatter histogram
 - Histograms show single-dimensional data of a cell population
 - Scatter plots show multi-dimensional data



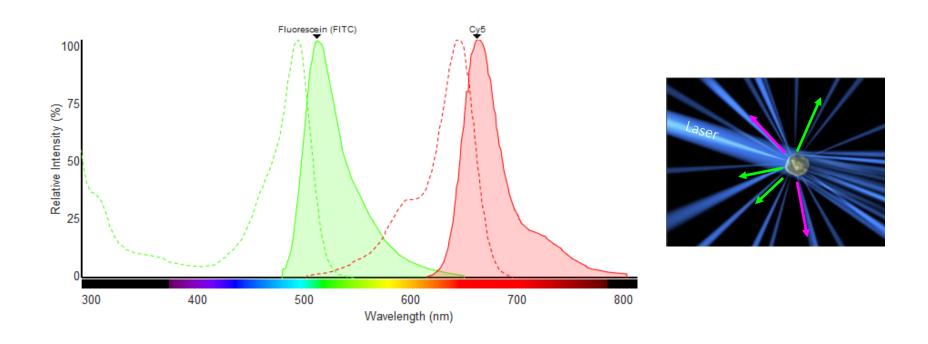


Whole blood scatter plot



Flow cytometry with fluorescence markers

- Cells can be labelled with fluorophore-coupled antibodies
- Fluorophores are chosen so as to have **distinct emission spectra** (e.g. 'green' and 'red' like FITC and Cy5)
- A cell expressing the corresponding antigen(s) will bind fluorophore-antibody conjugates and emit fluorescence upon passing through the laser beam.



Example: quantifying T cell populations from thymus

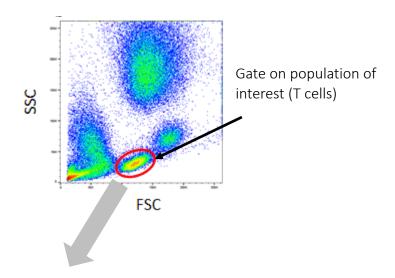
Label cells with antibodies

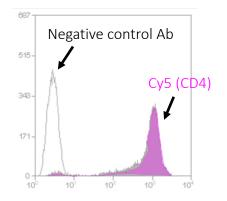
Quantify cell-associated fluorescence

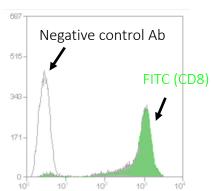
Plot cell counts

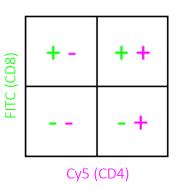
FITC-coupled anti-CD8 antibody CD4 antibody

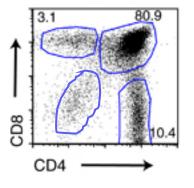




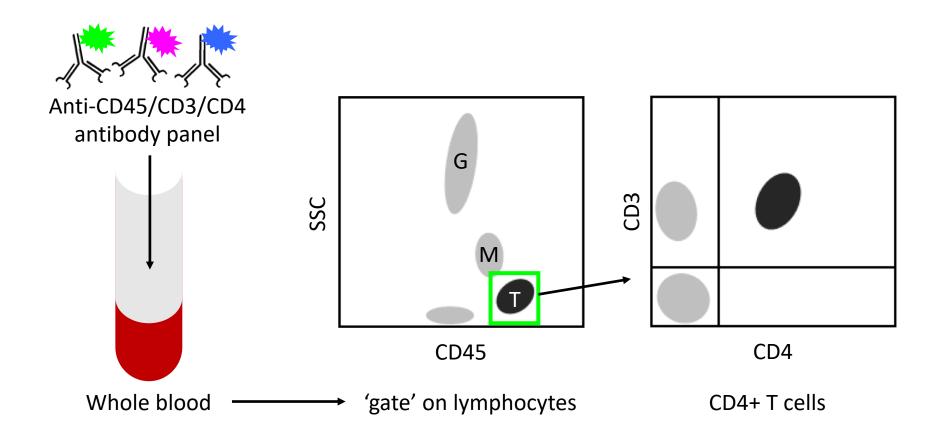








Example: CD4+ T cell counts in HIV infection



FACS (Fluorescence-Activated Cell Sorting)

- As cells pass through the laser beam, their characteristics are analysed
- Cells are given a charge based on their fluorescence profile
- Charged cells are deviated between deflector plates
- FACS yields highly purified cell populations
- Applications:
 - purify cells expressing a GFP-tagged protein
 - purify subpopulations of lymphocytes from blood
 - purify bone marrow stem cells for transplantation
 - ..

